Figure 1. Distinguishable and Indistinguishable Genotypes

When the wild-type and mutant homozygous genotypes are symmetric with respect to nearest-neighbor thermodynamics, their normalized melting curves are indistinguishable. This figure shows three replicate normalized melting curves of each of the three genotypes of the H63D (Hemochromatosis) mutation, which obeys this symmetry. Six curves representing the wild-type and mutant homozygous genotypes appear as one curve, while the normalized melting curves of the heterozygote appear as another easily distinguishable curve.

Figure 2. Theoretical duplex melting curves, superpositions and differences

After amplification of heterozygous DNA (or a mixture of any two genotypes of a biallelic SNP) two types of forward strands reassociate independently with two types of reverse strand, forming two species of homoduplex and two species of heteroduplex. A fluorescent dye gives a signal proportional to the quantity of DNA which is in its double-stranded state, which depends on the temperature and the thermodynamic stability of each duplex type. The overall fluorescence vs. temperature 'melting curve' is a superposition of the melting curves of the individual duplexes in proportion to their relative concentrations.

This figure gives the melting curves for the four duplex species of heterozygous or mixed genotype H63D amplicons as predicted by nearest-neighbor theory model with parameters described in (SL match and CC,GG). The wild-type and mutant homozygous curves are identical. It also shows the theoretical heterozygote melting curve obtained as the equally weighted average of the four duplex curves. In the nearest-neighbor symmetric case, the difference between any genotype mixture melting curves is proportional to a standard difference curve which is the difference between the average homoduplex curve and the average heteroduplex curve, by a factor equal to the absolute difference in heteroduplex content. Thus, the difference between the heterozygote melting curve and the overlapping homoduplex curves, which is also shown, is equal to the standard difference curve scaled by $\frac{1}{2}$, the heteroduplex fraction of an amplified heterozygote.

Figure 3. Theoretical Dependence Of Heteroduplex Content On Mixture Fraction

This figure shows the magnitude of the heteroduplex content differences among mixtures obtained by adding wild-type DNA to DNA of each of the three genotypes of a bi-allelic SNP (W=wild-type, M=mutant homozygous, H=heterozygous) in the relative proportions x of added wild-type (the 'mixture fraction') to 1 - x of the varied genotype. We assume that strands of different types associate independently after either PCR amplification is completed or a subsequent dissociation by heating and reassociation by cooling have been performed. We seek the mixture fraction providing the greatest separation of heteroduplex content between the resulting mixtures, represented by the highest point on the lowest among the three graphs. This point which is marked at $x = \frac{1}{7}$, gives the optimal mixture fraction.

Figure 4. Experimental Melting Curve Separation Dependence On Mixture Fraction

This figure shows the mean and standard deviation (error bars) of the experimentally measured maximum melting curve separation between replicates of mixtures of wild-type DNA with either mutant heterozygous or heterozygous samples, and the mean of the wild-type replicates, as a function of the mixture fraction. The values are normalized by a factor which makes the difference for an unmixed heterozygous sample equal to $\frac{1}{2}$ to agree with the theoretical prediction in terms of heteroduplex content (superimposed).

Figure 5a. Genotype Separation At Optimal Mixture Fraction 4/28

This figure shows the normalized melting curves for three replicates of optimal mixtures of wild-type DNA and each genotype of sample. As predicted, the mutant homozygous curves lie equidistant from the wild-type curves and the barely altered heterozygous curves. Figure 5b. Genotype Separation At Non-Optimal Mixture Fraction 9/28

Figure 5c. Genotype Separation At Non-Optimal Mixture Fraction 14/28

Figure 5d. Genotype Separation At Non-Optimal Mixture Fraction 19/28

These figures show the normalized melting curves for three replicates of other-thanoptimal mixtures of wild-type DNA and each genotype of sample. In Figure 5b, with mixture fraction is $\frac{9}{28}$ near $x = \frac{1}{3}$, the mutant homozygous curves and the heterozygous curves have become indistinguishable, as predicted. In Figure 5c, with mixture fraction is $\frac{14}{28}$, i.e., wild-type and sample DNA mixed in equal proportions), the mutant homozygous curves and the heterozygous curves are also difficult to distinguish. In Figure 5d, with mixture fraction is $\frac{19}{28}$ near the upper local maximum $x = \frac{2}{3}$, the mutant homozygous curves and the heterozygous curves have reversed position and are distinguishable, but not nearly as well as at the optimal mixture fraction.

Figure 6a. Experimental Difference Curves Of Pure Genotypes

Figure 6b. Experimental Difference Curve Of Optimally Mixed Genotypes

These figures show the difference of normalized melting curves of unmixed and optimally mixed samples of the three genotypes and the mean of the wild-type normalized melting curve replicates. The shape of the curves reflects the predicted standard difference curve shape, and the amplitude agrees with the predicted dependence on heteroduplex content difference.

Figure 7a. Raw TGCE Duplex Arrival Frequency Data

Figure 7b. Normalized TGCE Duplex Arrival Frequency Data

These figures show the raw and normalized measurements of duplex arrival frequency (counts per frame) for three replicates of each genotype and wild-type DNA with mixture fraction $x = \frac{9}{28}$. After normalization by shifting and scaling the largest peaks to a common location and height, the common heteroduplex content of the mutant homozygous and heterozygous sample mixtures for this mixture fraction are visible from the common first and second heteroduplex peak heights.

Figure 8. Heteroduplex Content Dependence On Mixture Fraction From TGCE

This figure shows the mean and standard deviation (error bars) of the experimentally derived heteroduplex content of replicates of mixtures of wild-type DNA with either mutant heterozygous or heterozygous samples, as a function of the mixture fraction. The values are adjusted by a factor which makes the value for control heterozygous samples equal to $\frac{1}{2}$ to agree with the theoretical prediction of heteroduplex content (superimposed).